

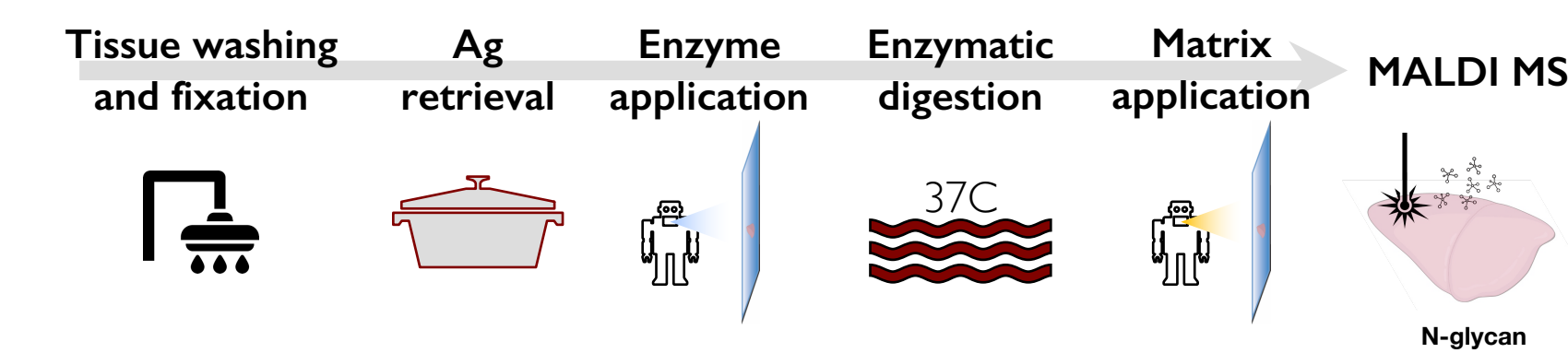
Single cell multi-omic mass spectrometry imaging for the human bone marrow microenvironment

Introduction

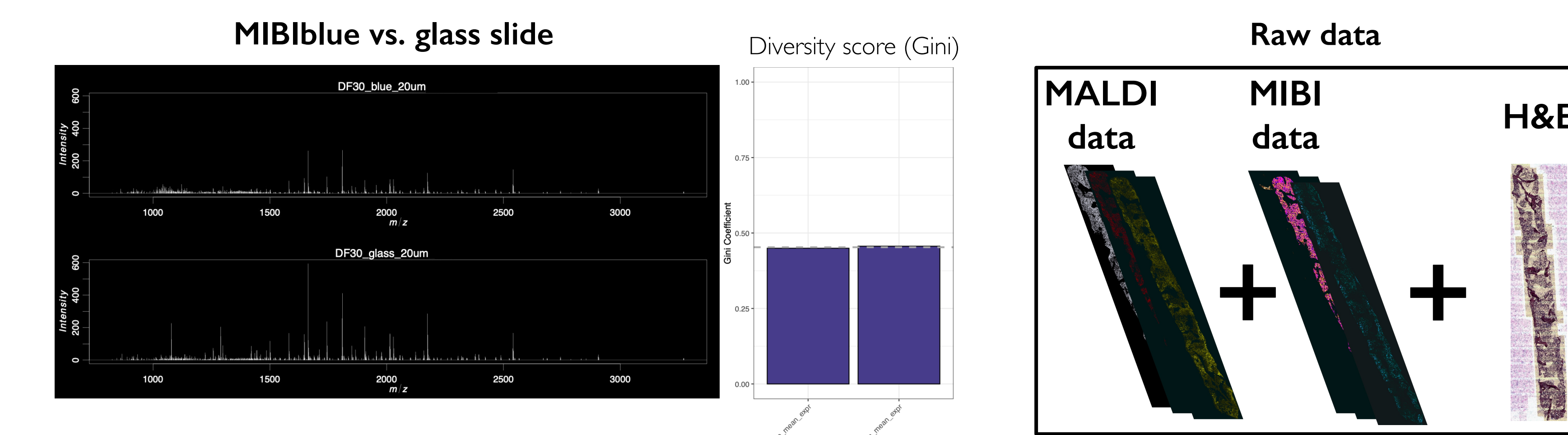
- The bone marrow structure reflects evolution-driven mechanisms that have tailored bone to create unique microenvironments catering to the diverse needs of developing blood cells.
- Glycosylation constitutes an essential organizing layer within this system, distinguishing itself from proteomic or transcriptomic properties.
- A scalable workflow has been developed for the multi-platform, sequential mass spectrometry-based imaging of bone marrow tissue sections.
- This workflow incorporates N-Glycan quantification through matrix-assisted laser desorption ionization (MALDI) followed by deep single-cell phenotyping via multiplex ion beam imaging (MIBI).
- The same tissue area is examined using both these methods, along with conventional, high-resolution, light-based Hematoxylin and Eosin (H&E) staining.

Analysis Strategy (see Poster WP377)

FFPE tissue sections were sectioned onto an organic polymer-coated slide (MIBIblue). The tissue was imaged with a MALDI timsTOFflex using a published protocol for N-glycan imaging (1).

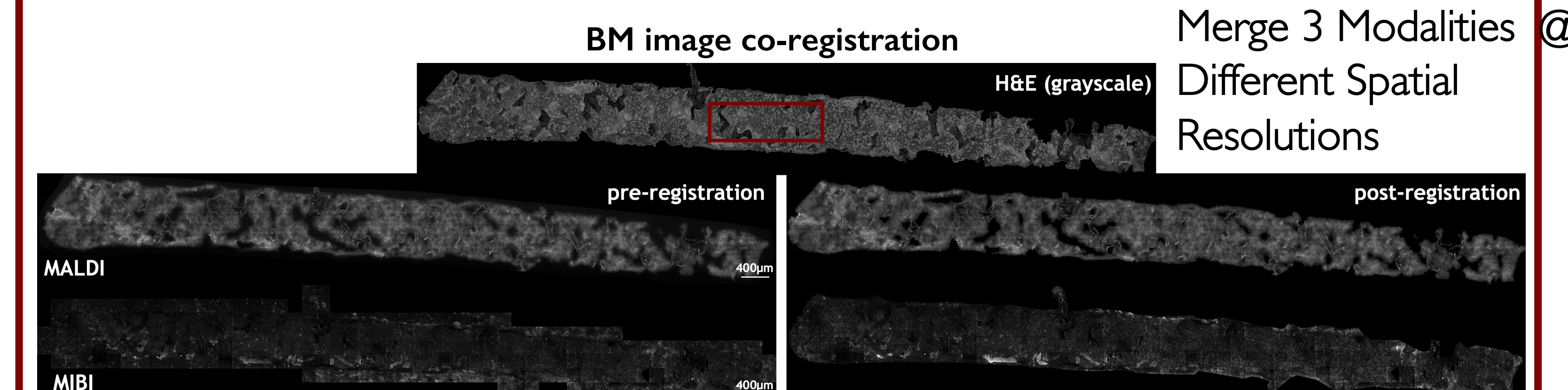


The matrix was washed off and sections were stained with a mix of metal-labelled antibodies targeted to immune phenotypic markers (2). Data was acquired with a MIBIScope and sections were counterstained with H&E.

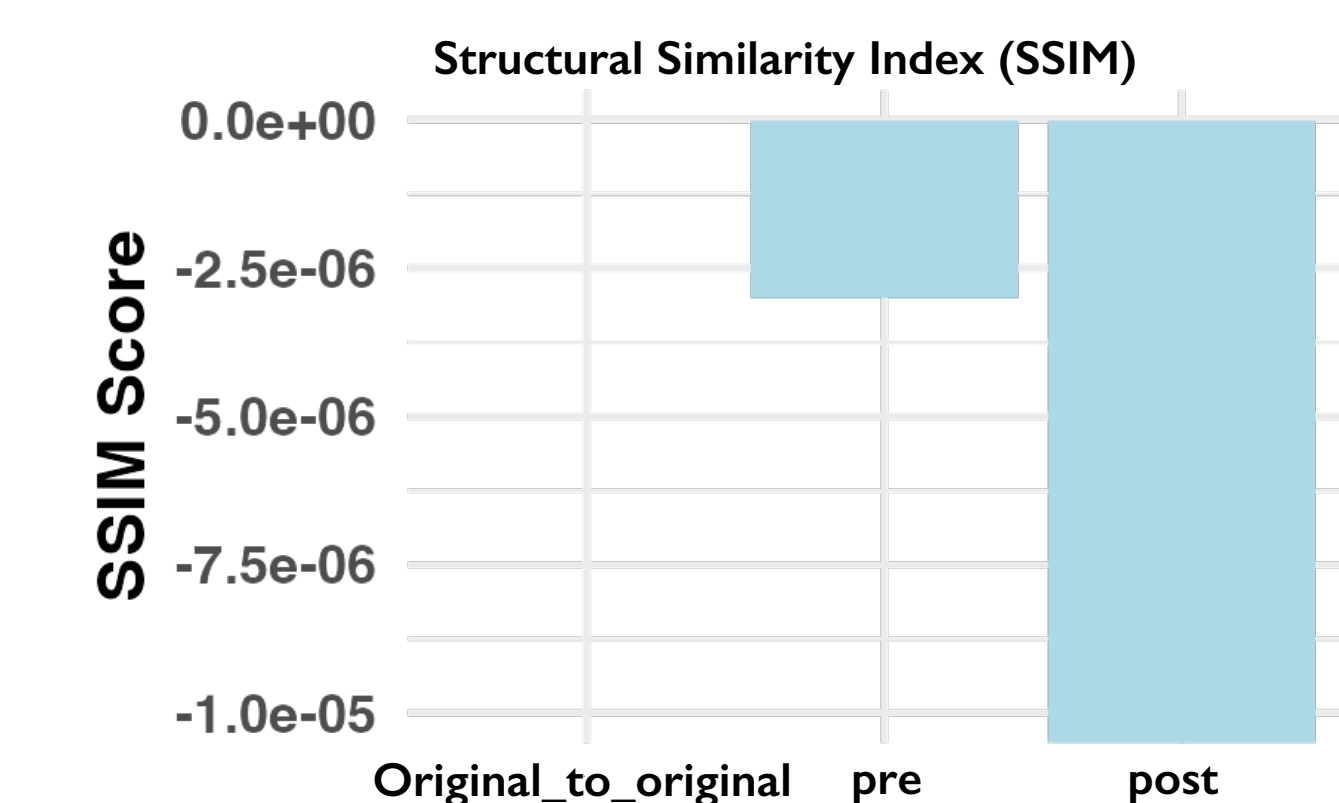


Integration of Cell Features & Glycans

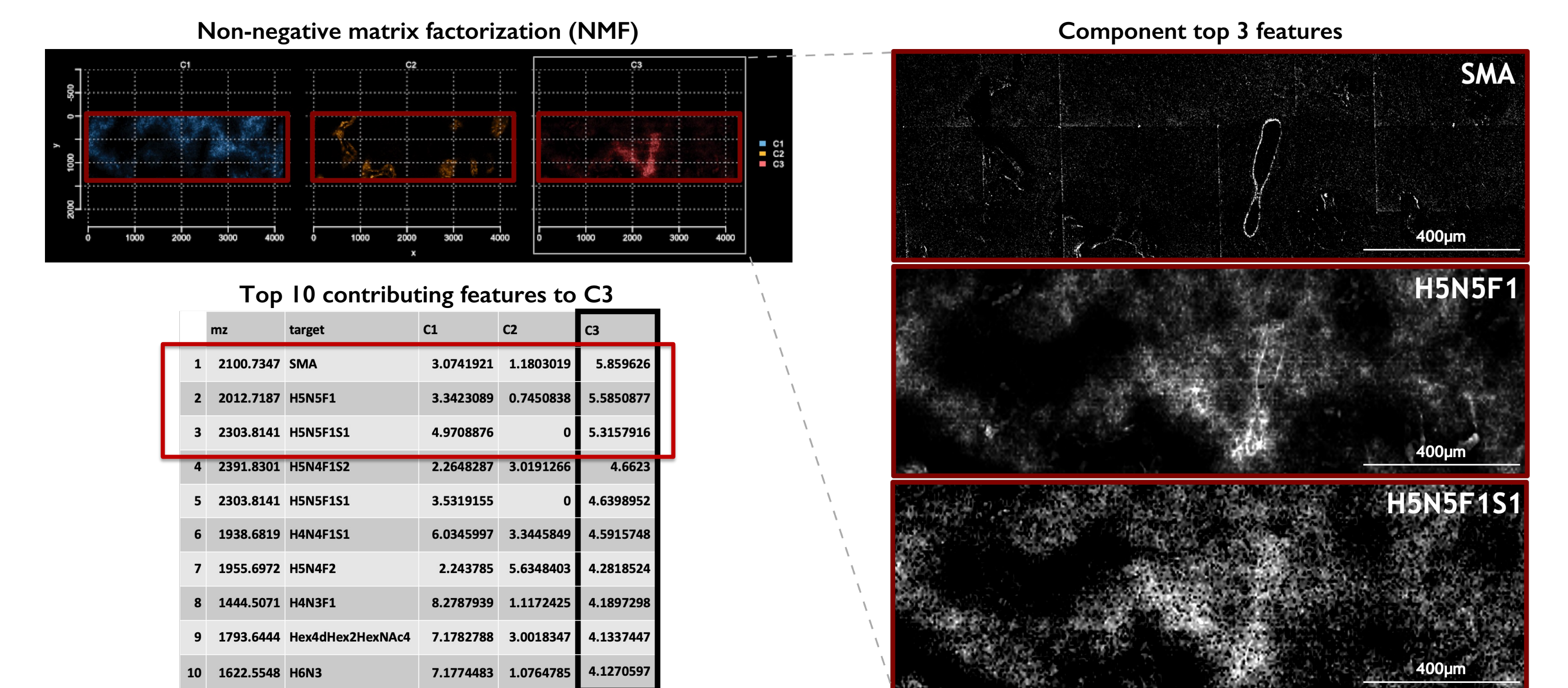
Registered MnM (MALDI & MIBI) imaging data were integrated into a single dataset where each pixel retains bimodal information from N-glycans and MIBI probes. SSIM scores indicates successful structural similarity as result of MALDI pixel up-sampling and landmark image registration to MIBI.



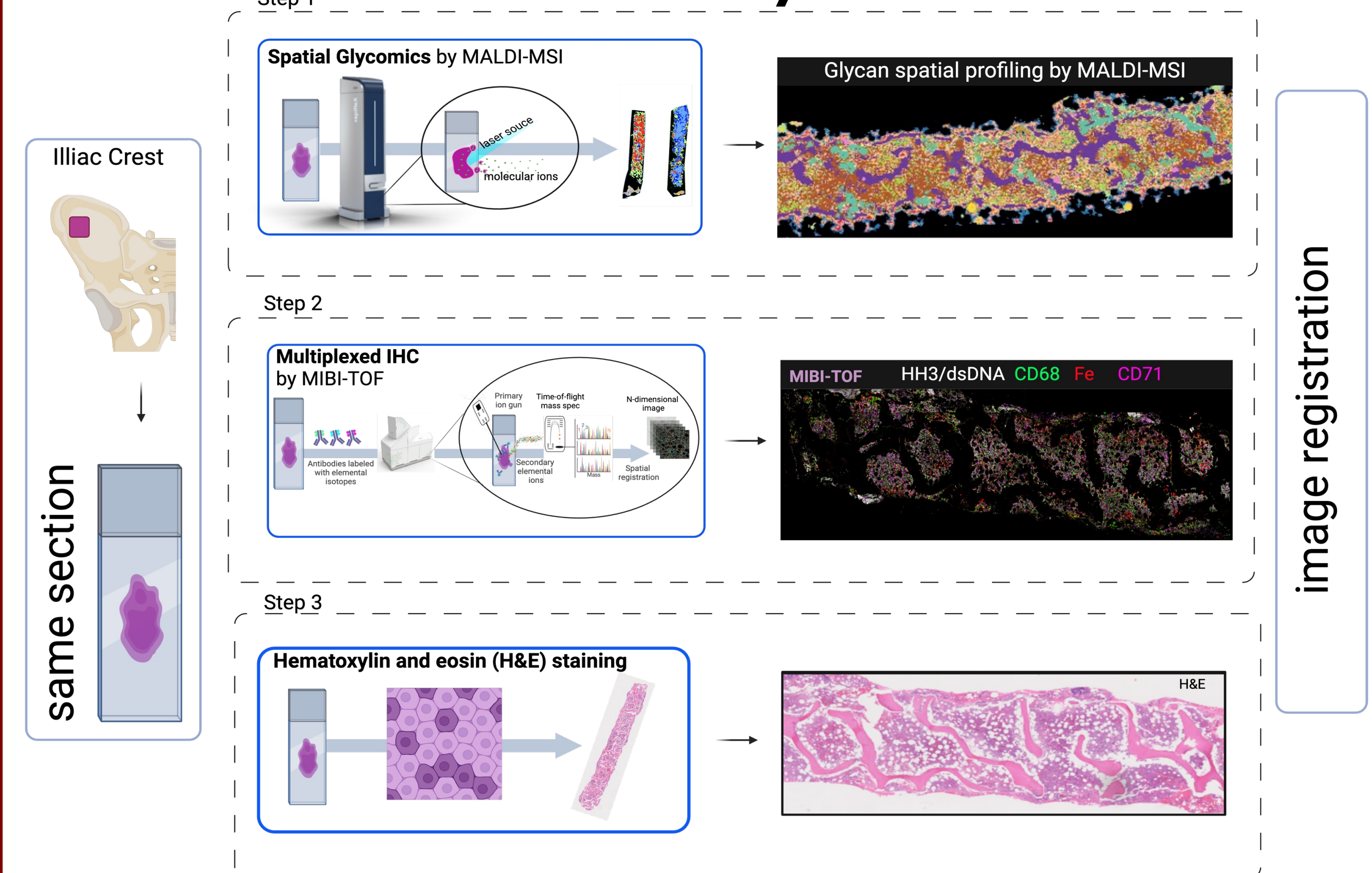
Resolution-based Merging Increases Spatial Similarity



Identify Top Molecular (Glycan) Components with sub micron-scale (Protein) Structures



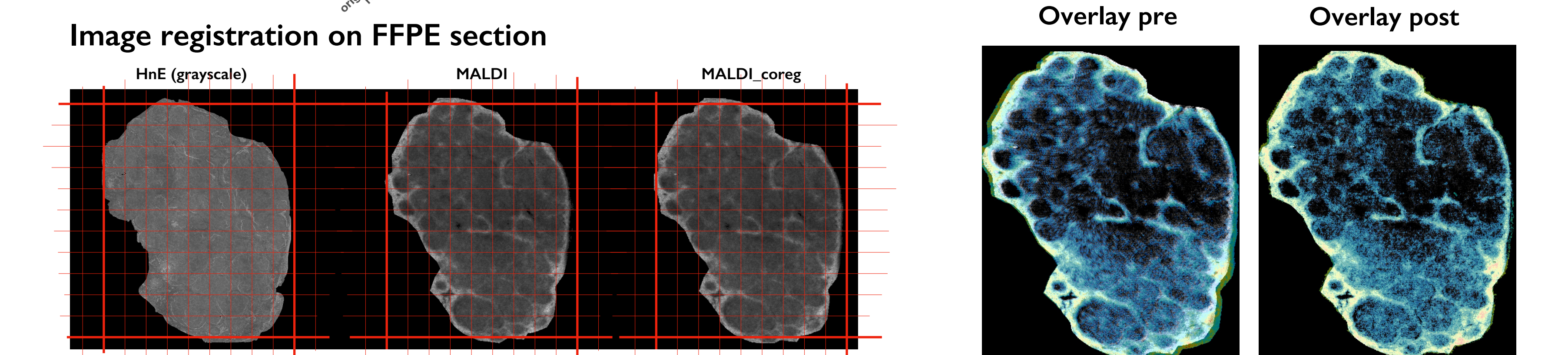
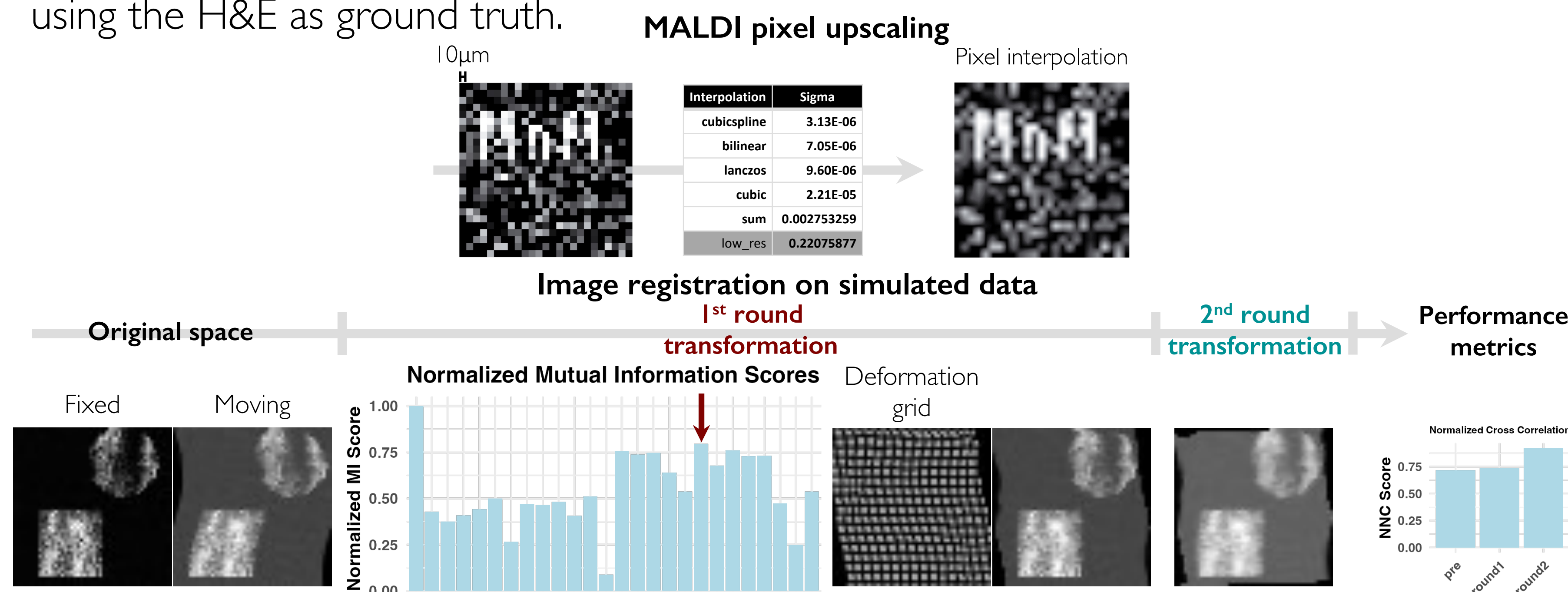
Multi-Omic Analysis Scheme



By imaging up to 40 proteins at the same time using MIBI-TOF, we can comprehensively identify at a *single-cell level*, the lineage, function, and spatial location of every cell in a single bone marrow tissue section. MIBI-TOF subcellular imaging will be used to capture the composition, functional state, and spatial organization of BM. These data will be complemented by a breakthrough de novo imaging that permits direct identification of glycans using matrix-assisted laser desorption ionization imaging (MALDI). The H&E stain is the final step, providing additional information about the cellular and tissue organization in the bone marrow.

Pixel resampling and iterative registration

Image registration methods were tested on simulated data. Bilinear interpolation was used for pixel up-sampling of the MALDI dataset to match the MIBI pixel size. Alignment of the two modalities into a shared coordinate system was achieved by using the H&E as ground truth.



Related Presentations

- Talk – Ke Leow - Wed Jun 5 – 8:30am (WOA Room 210ABC) – Spatial Multi-omics (Single Cell Proteomics, Spatial Glycan, Spatial Trxn) in Human Pregnancy
- Poster – Mikaela Ribí – Mon Jun 3 - MP 291 – MALDI N-Glycomics and High-Grade Glioma
- Poster – Davide Frachina – Wed Jun 5 – WP 377 – Co-Spatial Tissue Imaging (Single cell Proteomics, N-Glycomics, H&E bright field)

References

- Drake RR *et al.* In Situ Imaging of N-Glycans by MALDI Imaging Mass Spectrometry of Fresh or Formalin-Fixed Paraffin-Embedded Tissue. 2018
- Liu CC *et al.* Reproducible, high-dimensional imaging in archival human tissue by multiplexed ion beam imaging by time-of-flight (MIBI-TOF). 2022

Disclosure: SCB and MA are founders of Ionpath which commercializes MIBI technology; work funded by NIH U54HL165445, R01AG078702, U24CA224309

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